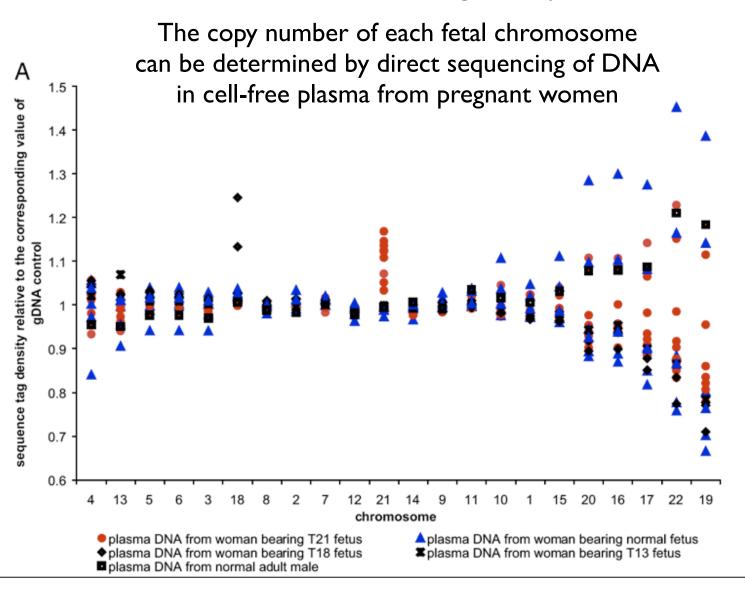
#### In the news:

# Noninvasive diagnosis of fetal aneuploidy by shotgun sequencing DNA from maternal blood

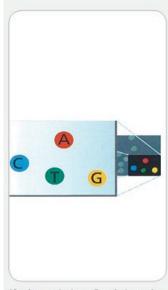
H. Christina Fan\*, Yair J. Blumenfeld<sup>†</sup>, Usha Chitkara<sup>†</sup>, Louanne Hudgins<sup>‡</sup>, and Stephen R. Quake\*<sup>§</sup>



# 7. DETERMINE FIRST BASE

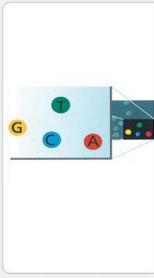
First chemistry cycle: to initiate the first sequencing cycle, add all four labeled reversible terminators, primers and DNA polymerase enzyme to the flow cell.

#### 10. IMAGE SECOND CHEMISTRY CYCLE



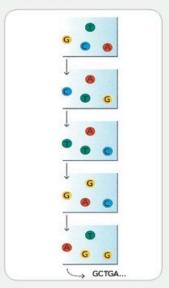
After laser excitation, collect the image data as before. Record the identity of the second base for each duster.

## 8. IMAGE FIRST BASE



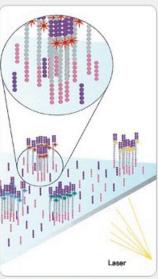
After laser excitation, capture the image of emitted fluorescence from each duster on the flow cell. Record the identity of the first base for each duster.

#### 11. SEQUENCE READS OVER MULTIPLE CHEMISTRY CYCLES



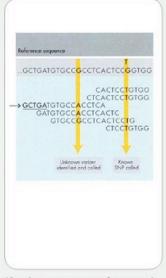
Repeat cycles of sequencing to determine the sequence of bases in a given fragment a single base at time.

#### 9. DETERMINE SECOND BASE



Second chemistry cycle: to initiate the next sequencing cycle, add all four labeled reversible terminators and enzyme to the flow cell.

#### 12. ALIGN DATA



Align data, compare to a reference, and identify sequence differences.

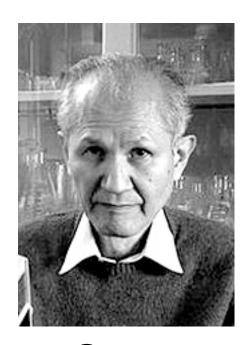
## High-throughput sequencing using Solexa/Illumina technology

#### **Confession**:

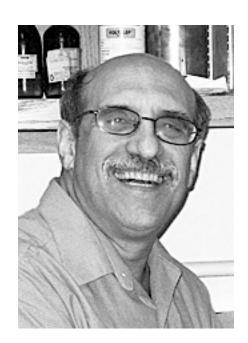
I think I'm one of those people who's happy to use highthroughput sequencing without a complete understanding of how it works.



## Today's Nobel Prize in Chemistry



Osamu Shimomura



Marty Chalfie



Roger Tsien

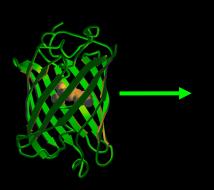
## Green Fluorescent Protein (GFP)

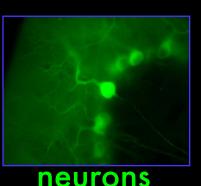
Comes from a jellyfish, Aequorea victoria

Gene has been cloned and transferred into a wide variety of "heterologous" expression systems

... including Drosophila, mammalian cells, C. elegans, yeast, zebrafish etc. etc.

\*\*\*\* Permits dynamic and in vivo analysis\*\*\*\*
of biological processes

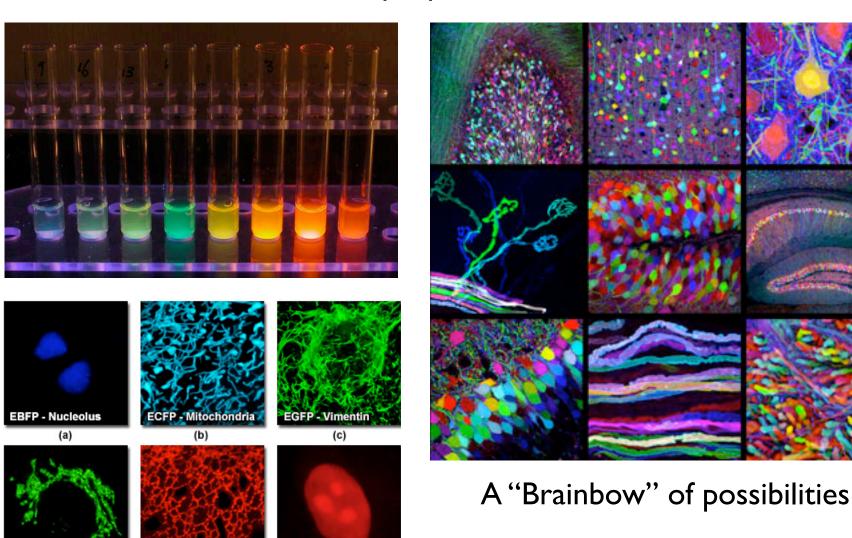




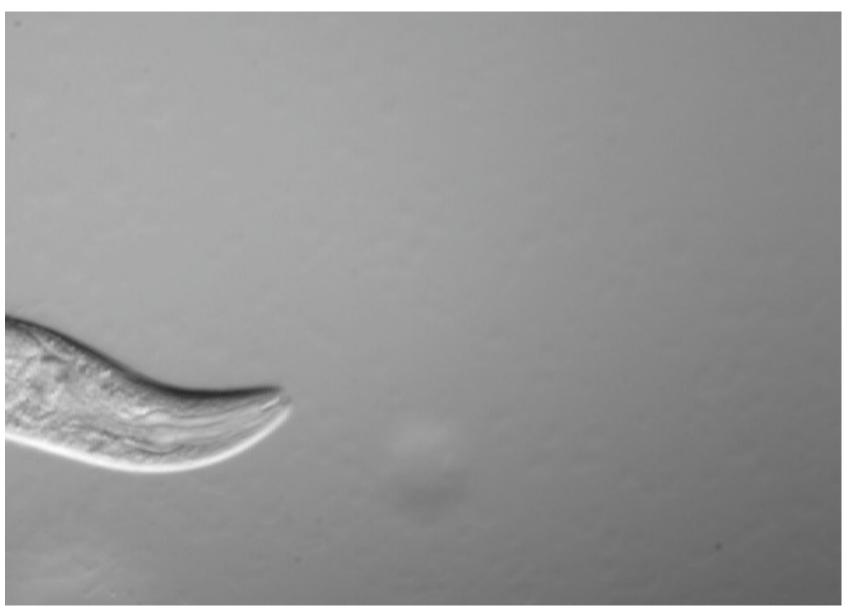




Variants of Green Fluorescent Protein and DsRed have been engineered to have different excitation and emission spectra, and other useful properties



## It's a bird! It's a plane! It's.... C. elegans!



Laboratory of Bob Goldstein, UNC

# Reading: the Portrait chapter (will be posted on the course website today)

#### Caenorhabditis elegans: Genetic Portrait of a Simple Multicellular Animal



The nematode Caenorhabditis elegans, one of the simplest multicellular organisms, lives in soils worldwide and feeds on soil bacteria. Adults are about 1 mm in length and contain an invariant number of somatic cells (Fig. C.1). The mature "female," which is actually a hermaphrodite able to produce both eggs and sperm, has precisely 959 somatic cells that arose from progenitor cells by a reproducible pattern of cell division. The mature male, which produces sperm and has genitalia that enable it to mate with the hermaphrodite, includes precisely 1031 somatic cells that also arose by a reproducible pattern of cell division. C. elegans has a short life cycle and an enormous reproductive capacity, progressing in just three days from the fertilized egg of one generation to between 250 and 1000 fertilized eggs of the next generation. It is transparent at all stages, so that investigators can use the light microscope to track development at the cellular level throughout the life cycle. Its small size and small cell number, precisely reproducible and

viewable cellular composition, short life cycle, and capacity for prolific reproduction make C. elegans an ideal subject for the genetic analysis of development. The fact



An adult C. elegans hermaphrodite surrounded by larvae of various



Sidney Brenner

Using C. elegans as a genetic model system was this guy's idea



John Sulston

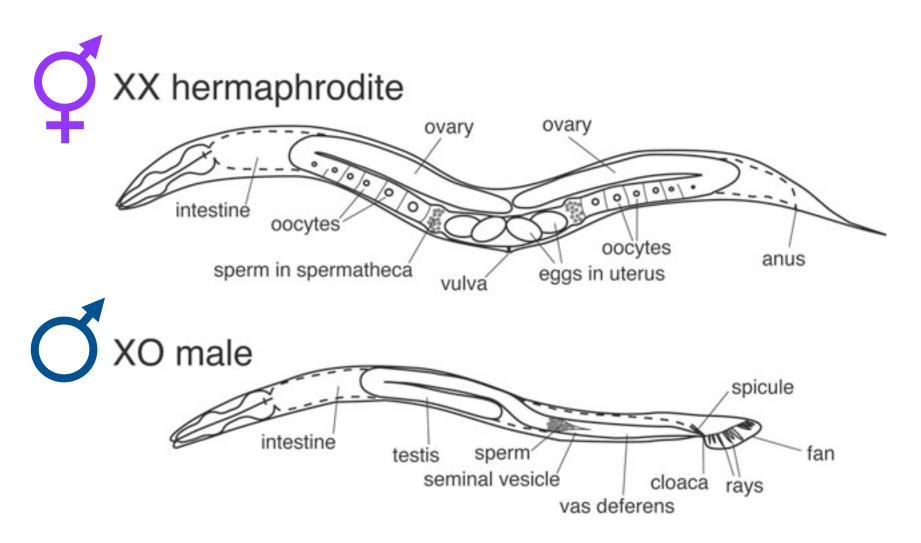
He shared the 2002 Nobel prize with these guys for working out the cell lineage and apoptosis



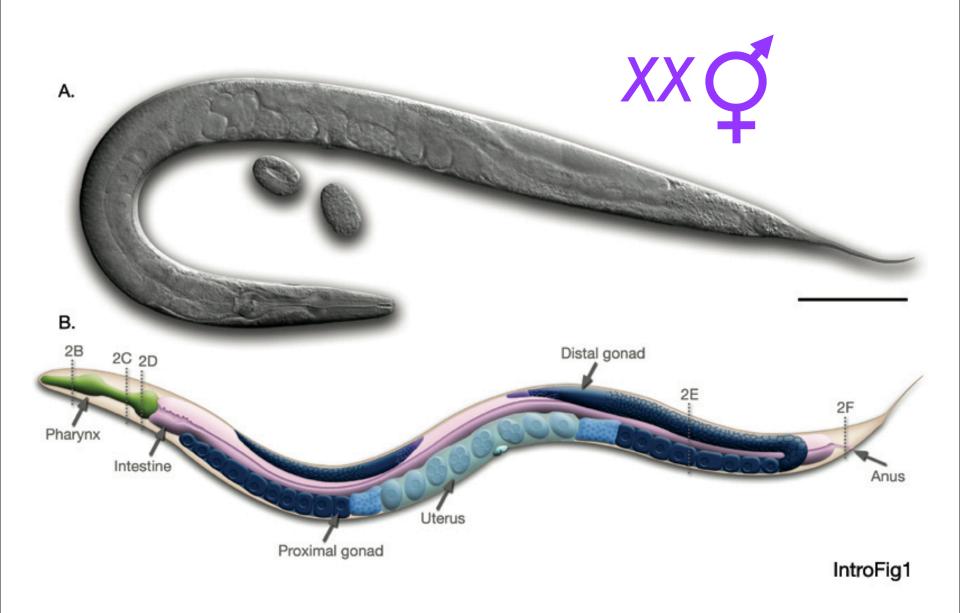
**Bob Horvitz** 



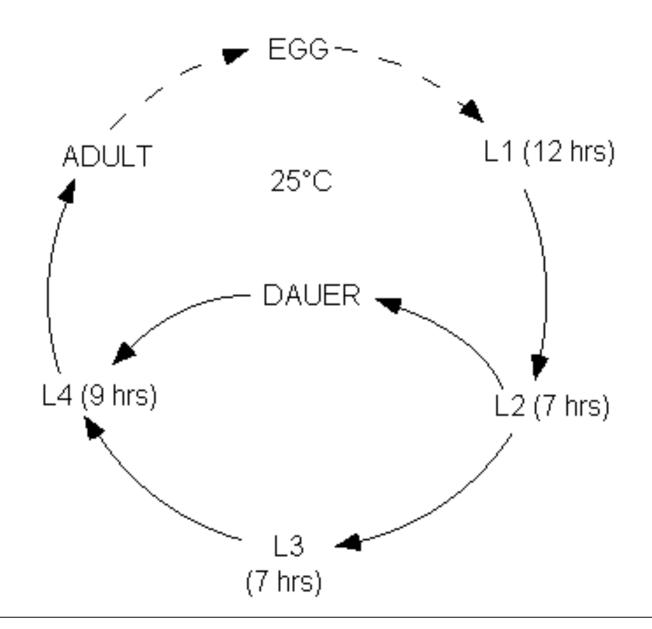
# Anatomy of the worm



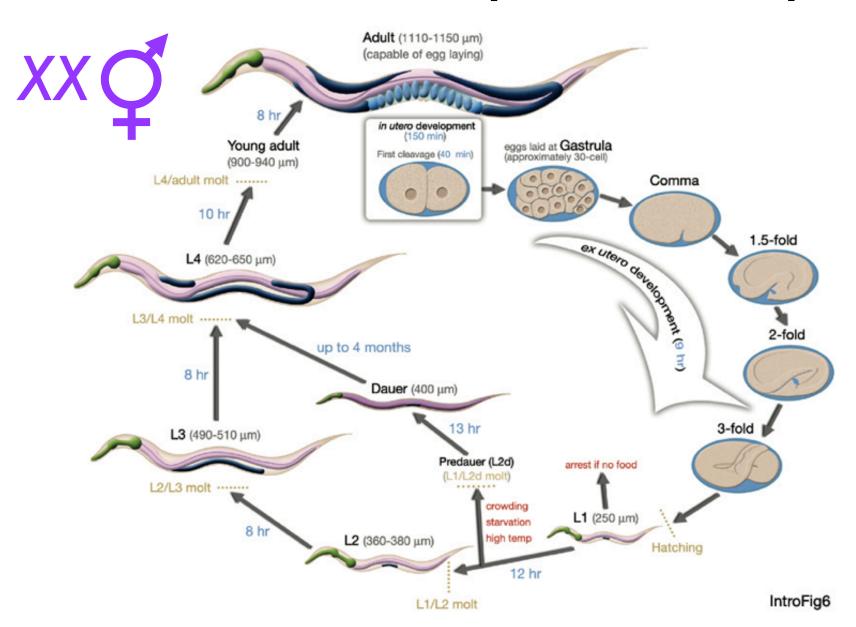
## Anatomy of the worm



# C. elegans Development

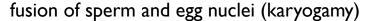


# The worm life cycle: 3.5 days

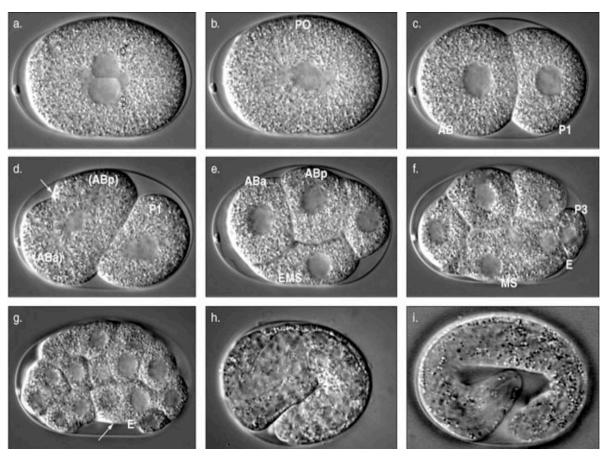


## Embryonic development

(takes ~24 hrs at 20°C)



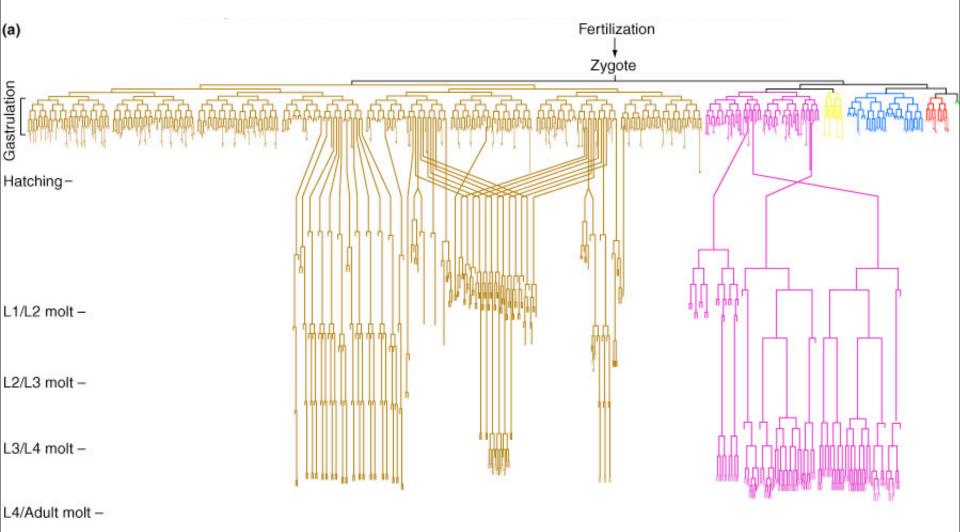
I<sup>rst</sup> mitotic division



"comma stage" embryo

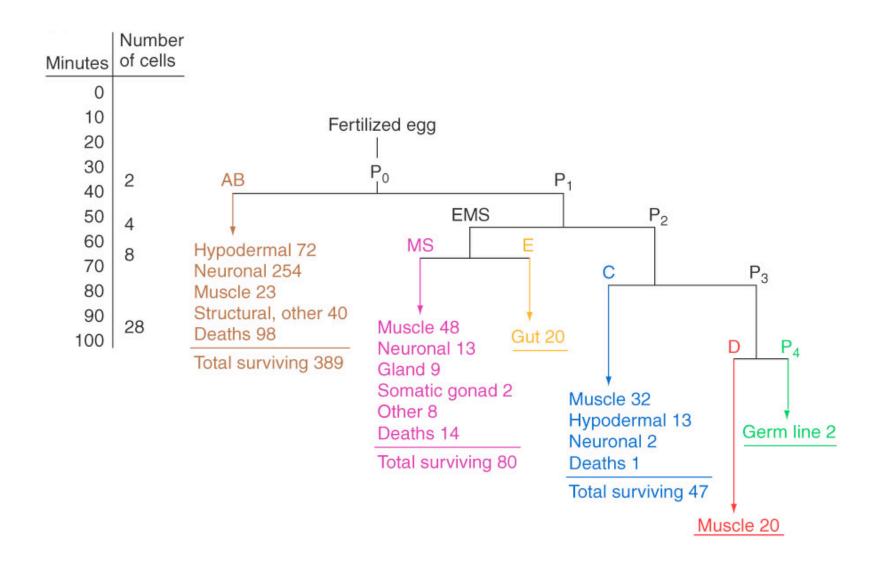
"pretzel stage" embryo (about to hatch as LI larva)

## C. elegans has an "invariant" cell lineage\*

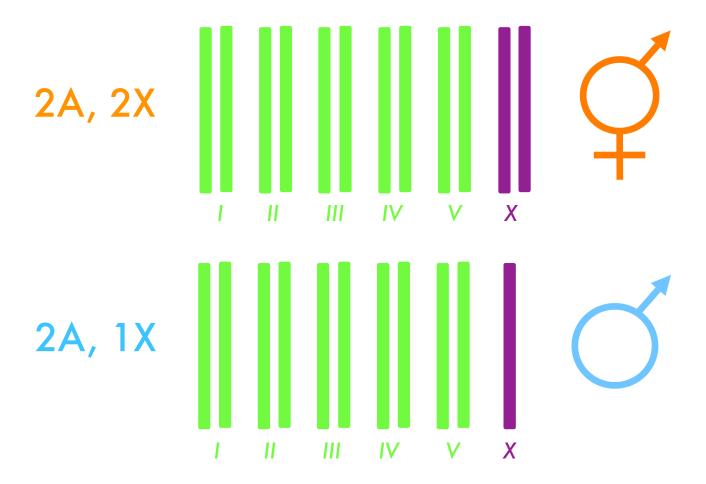


\*No, you do not have to memorize it.

# The earliest divisions give rise to many different tissues

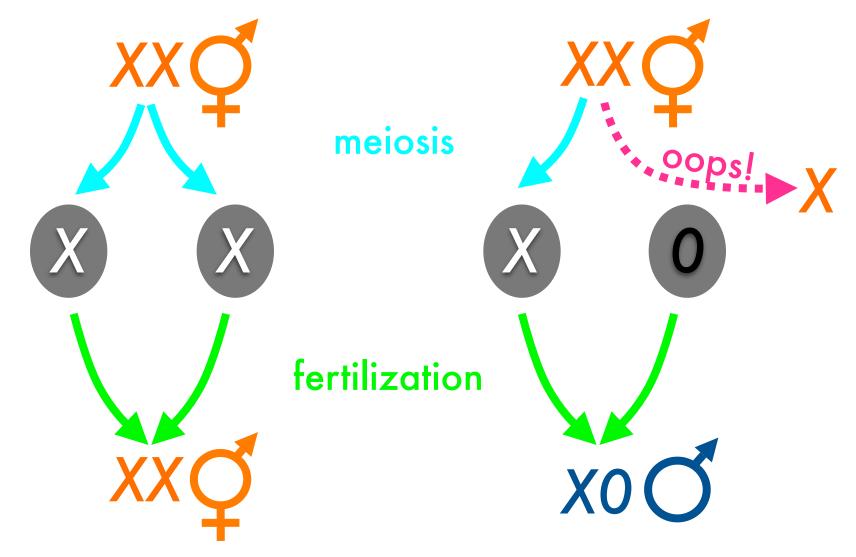


## Sex determination in C. elegans: XX and XO



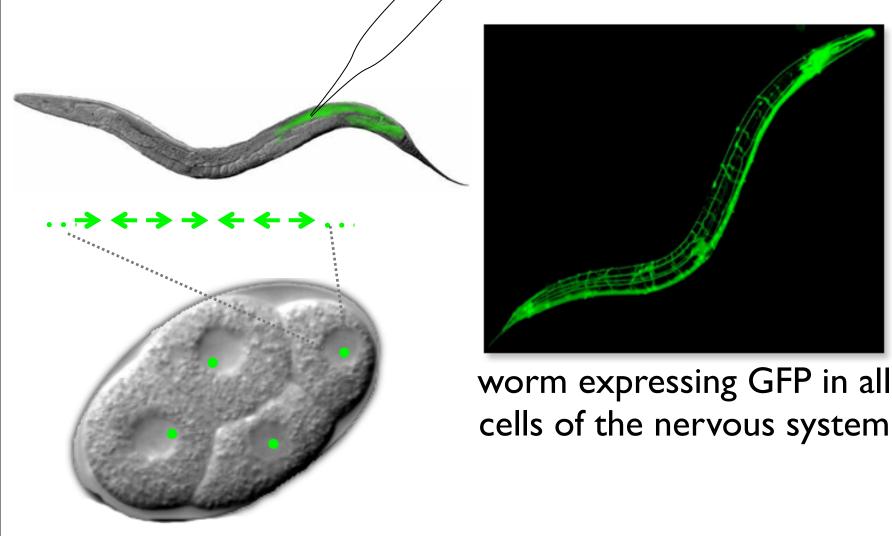
The chromosomes are drawn this way because they are "holocentric" (centromeres are distributed throughout). This is confusing at first when you think about meiosis, but you get used to it.

## Where do C. elegans males come from?



Hodgkin, Horvitz, and Brenner (1979), Genetics 91:67-94

Transgenic C. elegans can be made by injecting DNA into the gonad; some of the progeny will carry the genes that are injected, in high copy number



A basic screen for recessive mutations in C. elegans generation ethyl methanesulfonate  $P_0$ makes mostly G->A mutations self-fertilize heterozygous  $\mathsf{F}_{\mathsf{I}}$ for any new mutation self-fertilize total elapsed time: ~I week.

# Worms are simple creatures, and so many mutations cause the same general phenotype

Unc = Uncoordinated (aberrant or absent movement)

Dpy = Dumpy (short and/or fat)
(can result from hyperexpression of the X chromosome)

Let = Lethal

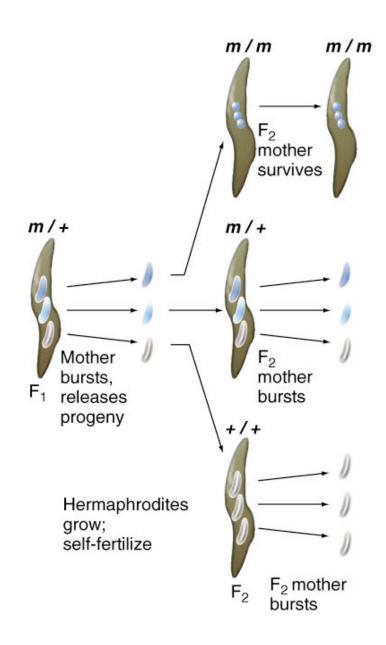
Emb = Embryonic lethal (also Zyg, for zygotic lethal)

Lon = Long and thin

Phenotypes are Capitalized (Unc), genes are *lower-case and italicised*, with 3 letters, a hyphen, and a number (*unc-51*), and the encoded proteins are ALL CAPS (UNC-51)

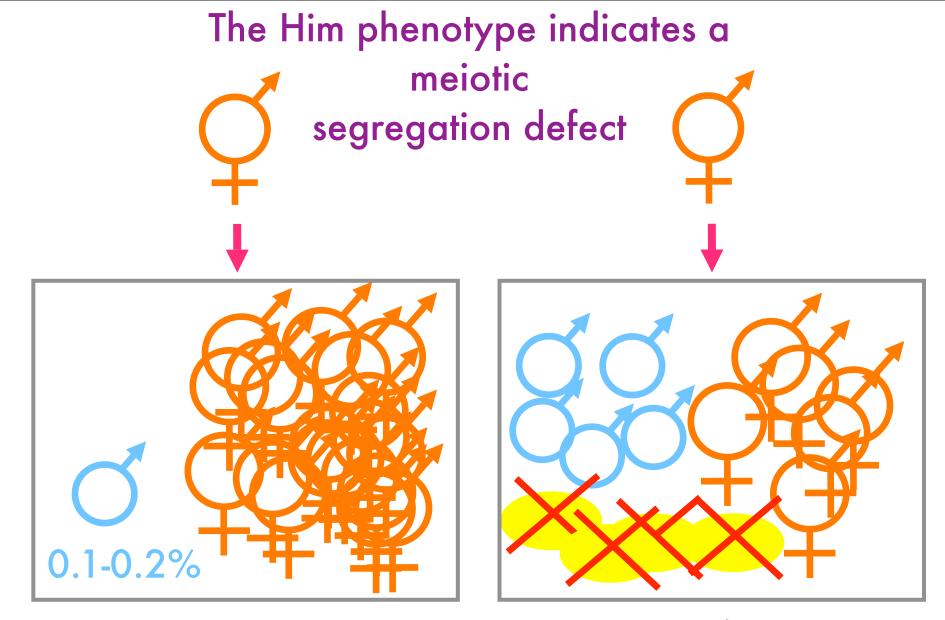


The "bag-of-worms" phenotype results from an inability to lay eggs (Egl - egg laying defective or Vul - vulvaless)



This (admittedly gross)
phenomenon can be used to
screen for "maternal effect
lethals" (Mel mutants:
homozygous mothers are
o.k., but their embyros die.

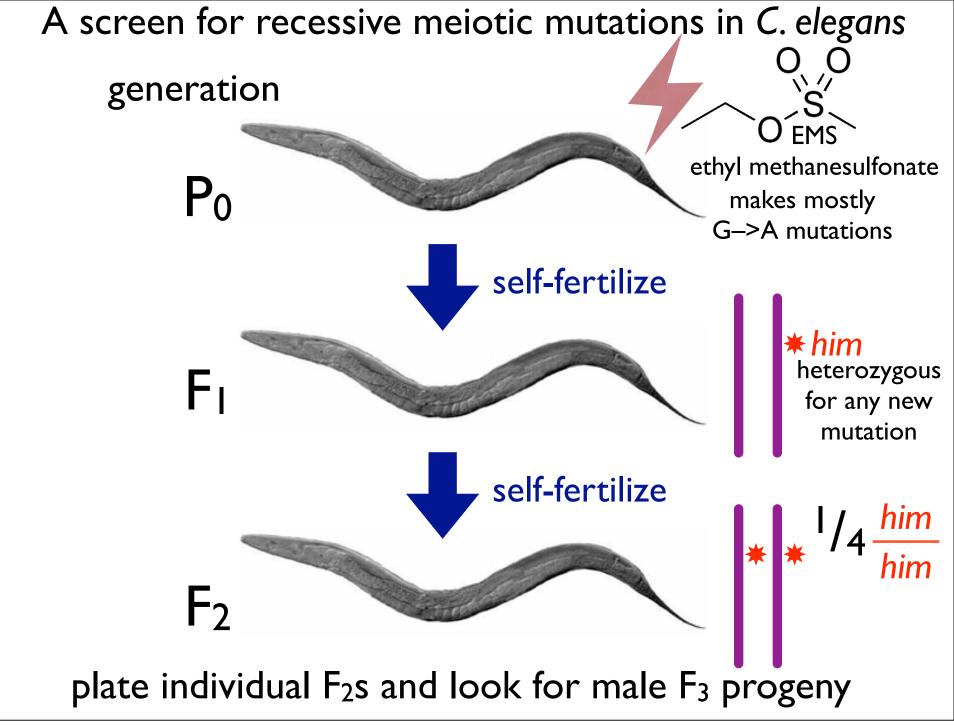
One class of Mel mutants are severely defective in meiosis - they produce aneuploid embryos, which die.



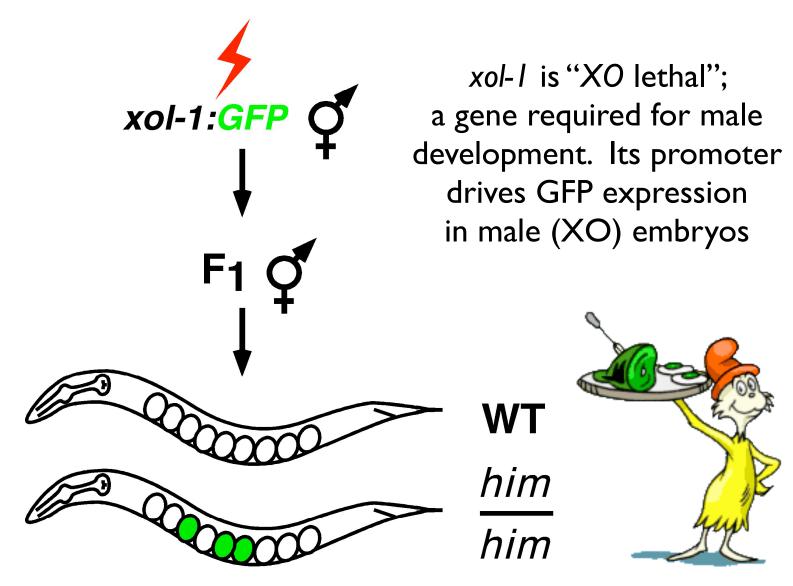
normal hermaphrodite

High incidence of males (Him)

Hodgkin, Horvitz, and Brenner (1979) Genetics 91: 67-94

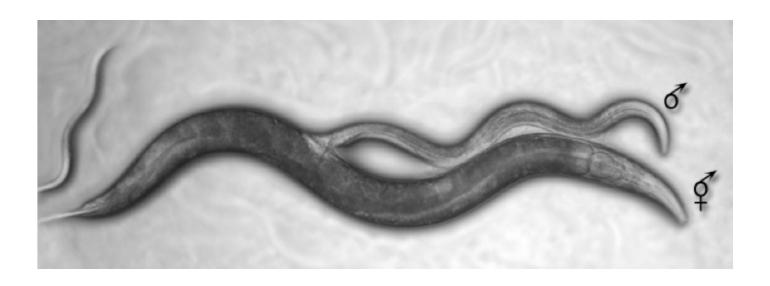


A simpler way to screen for meiotic mutants: look for the Him phenotype using the "Green Eggs and Him" trick





## Anatomy of the worm



mating

Note: Mating requires a lot of activity on the part of the male, but is essentially a passive process from the perspective of the hermaphrodite... this means that some mutations (like Unc mutations, which compromise mobility) cannot be homozygous/hemizygous in the male.

So how do you tell the difference between self and cross progeny?

Dpy 
$$(dpy-5 I) \neq x WT \checkmark$$

**|** 

self progeny

Dpy 🗸 (plus the rare 🗸)

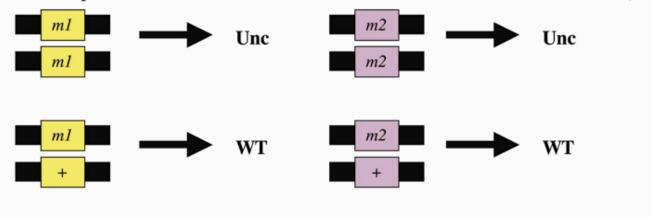
OR

cross-progeny 50% nonDpy ♂; 50% nonDpy ♂

Note: this example uses an autosomal dpy mutation. What would you expect if the dpy gene were on the X chromosome?

### Complementation tests in C. elegans are straightforward

m1 and m2 are two separate recessive mutations that both result in the same uncoordinated (Unc) behavior.



If both of these mutations are present in a trans configuration, and an uncoordinated behavior is observed



then these mutations DO NOT complement each other, and are alleles of the same gene.

But, if the *trans* configuration results in wild-type (WT) behavior



then these mutations DO complement each other, and are alleles of different genes.

Note: failure to complement usually, but not always means that mutations affect the same gene. Conversely, complementation usually but not always means that mutations are in different genes.

Next lecture (Friday):

mapping genes in *C. elegans*pathway analysis
how we sort out what meiotic genes do